

# Neutral Network Models for Biochemical Systems

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## Overview

### Background

The major difficulty in modeling biological systems from multivariate time series is the identification of parameter sets that endow a model with dynamical behaviors sufficiently similar to the experimental data. Directly related to this parameter estimation issue is the task of identifying the structure and regulation of ill-characterized systems.

### Results

We propose a method for the identification of admissible parameter sets of canonical S-systems from biological time series. The method is based on a Monte Carlo process that is combined with an improved version of our previous parameter optimization algorithm [3]. The method maps the parameter space into the network space, which characterizes the connectivity among components, by creating an ensemble of decoupled S-system models that imitate the dynamical behavior of the time series with sufficient accuracy. The concept of sloppiness is revisited in the context of these S-system models with an exploration not only of different parameter sets that produce similar dynamical behaviors but also different network topologies that yield dynamical similarity.

### Conclusions

The proposed parameter estimation methodology was applied to actual time series data from the glycolytic pathway of the bacterium *Lactococcus lactis* [2] and led to ensembles of models with different network topologies. In parallel, the parameter optimization algorithm was applied to the same dynamical data upon imposing a pre-specified network topology derived from prior biological knowledge, and the results from both strategies were compared. The results suggest that the proposed method may serve as a powerful exploration tool for testing hypotheses and the design of new experiments.

## Methods

$$S_i = \alpha_i \prod_{j=1}^n X_j^{\beta_{ij}} - \beta_i \prod_{j=1}^n X_j^{\gamma_{ij}} \quad \text{S-system equations [1]}$$

$$L = \begin{bmatrix} \beta_{11} \alpha_1 (\beta_{11}) & \dots & \beta_{1n} \alpha_1 (\beta_{1n}) & \dots & \beta_{1n} \alpha_1 (\beta_{1n}) \\ \beta_{21} \alpha_2 (\beta_{21}) & \dots & \beta_{2n} \alpha_2 (\beta_{2n}) & \dots & \beta_{2n} \alpha_2 (\beta_{2n}) \\ \vdots & \dots & \vdots & \dots & \vdots \\ \beta_{n1} \alpha_n (\beta_{n1}) & \dots & \beta_{nn} \alpha_n (\beta_{nn}) & \dots & \beta_{nn} \alpha_n (\beta_{nn}) \end{bmatrix} \quad \psi = \begin{bmatrix} \alpha_1 \\ \alpha_2 \\ \vdots \\ \alpha_n \end{bmatrix}$$

$$ES \text{ as Eigen space of } W \quad \begin{cases} \alpha_i \prod_{j=1}^n X_j^{\beta_{ij}} = ES \psi \\ \beta_i \prod_{j=1}^n X_j^{\gamma_{ij}} = ES \delta \end{cases} \quad S_i = \alpha_i \prod_{j=1}^n X_j^{\beta_{ij}} - \beta_i \prod_{j=1}^n X_j^{\gamma_{ij}}$$

$$\begin{cases} \alpha_i \prod_{j=1}^n X_j^{\beta_{ij}} = ES \psi \\ \beta_i \prod_{j=1}^n X_j^{\gamma_{ij}} = ES \delta \end{cases} \quad \begin{cases} \alpha_i \prod_{j=1}^n X_j^{\beta_{ij}} = e^{ES \psi} \\ \beta_i \prod_{j=1}^n X_j^{\gamma_{ij}} = e^{ES \delta} \end{cases} \quad S_i = \alpha_i \prod_{j=1}^n X_j^{\beta_{ij}} - \beta_i \prod_{j=1}^n X_j^{\gamma_{ij}}$$

$$F = \log \left( (S_i - \hat{S}_i(\delta)) / (S_i - \hat{S}_i(\psi)) \right)$$

$$\frac{\partial F}{\partial \delta} = ES \psi \left( \left[ (S_i + e^{ES \delta})^{-1} \cdot e^{ES \delta} \right] \delta ES \right)$$

$$\frac{\partial F}{\partial \delta} = 2 \left( -e^{ES \psi} \left( \frac{\partial S_i}{\partial \delta} \right) + e^{ES \delta} \delta ES \right) \left( S_i - \hat{S}_i(\delta) \right)$$

Leads to Levenberg-Marquardt algorithm

## Results

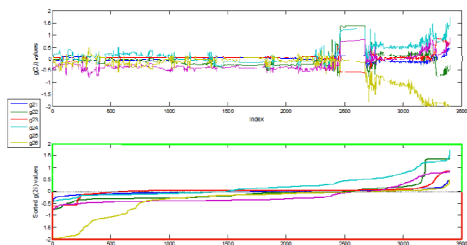


Figure 1 – Kinetic orders estimated for the G6P (X2) production term with potential inclusion of all system variables. A) The two indices refer to the two species considered by the interaction; for instance, g26 indicates the effect of X6 (acetate) on G6P production. B) The same parameter sets as shown in A, but ordered individually by magnitude, showing the possible variation admissible each parameter. The light green y-axis represents the region of parameter space with possible activation interaction; analogously, the light red y-axis represents the region of possible inhibition interaction.

Figure 2 – Quantification of sloppiness for one of the equations of the *Lactococcus* model. The figure shows the 3-D projection of the ellipsoid that represents the region of the parameter space that produces similar dynamical behaviors. The arrows show the direction of the sloppy and stiff directions in the 3-D projection, corresponding respectively to eigenvectors with small and large eigenvalues of the Hessian matrix of the cost function.

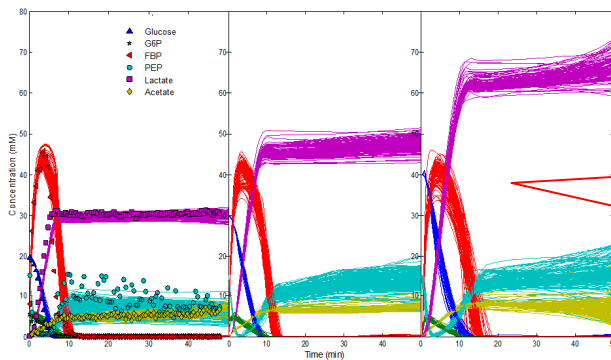
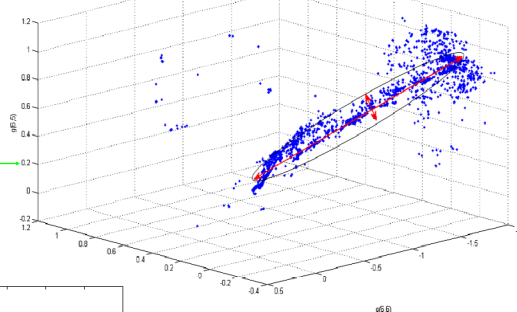
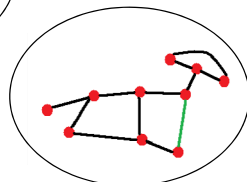
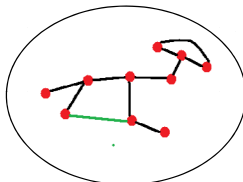
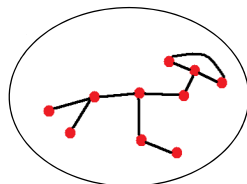
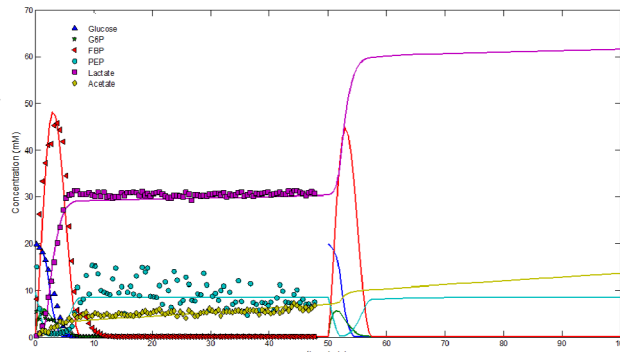


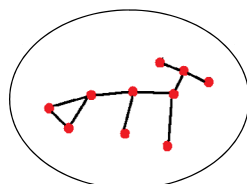
Figure 5 – Glucose double pulse simulation. A second 20 mM glucose pulse was supplied to the system after 50 min, resulting in the further accumulation of lactate and acetate.

Good generalization capability

New ensemble of models. Systems integrated with different initial concentrations for glucose substrate (20, 30 and 40 mM)



Neutral network space [4]



Robustness

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